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PATENT

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of: Riechmann, Lutz et al.

U.S. Serial No.: Not Assigned Yet

Filed: Concurrent Herewith

Entitled: "SELECTION SYSTEM"

Attorney Docket No.: 8654/1090

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09/710444
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Commissioner for Patents and Trademarks
Box Patent Application
Washington, D.C. 20231

TRANSMITTAL LETTER

Enclosed for filing in the above-identified patent application, please find the following documents:


1. Certificate of Express Mailing;
2. New Patent Application Transmittal;
3. Patent Application Specification, (53 pages); Claims (4 pages);
4. (6) Sheets of Informal Drawings;
5. Unexecuted Declaration and Power of Attorney;
6. Check for \$535.00 for the Requisite Filing Fee; and
7. Return Post Card.

Pursuant to 37 C.M.R. s.1.27, Applicant claims small entity status.

The Assistant Commissioner for Patents is hereby authorized to charge any additional fees or credit any overpayment in the total fees to Deposit Account No. 16-0085, Reference No. 8654/1090. A duplicate of this transmittal letter is enclosed for this purpose.

Respectfully submitted,

11/10/00
Date


Kathleen M. Williams, Esq.
Reg. No. 34,380
PALMER AND DODGE, LLP
One Beacon Street
Boston, MA 02109-3190
Telephone No: (617) 573-0451
Fax No: (617) 227-4420

09/710444-111000

NEW UNITED STATES UTILITY PATENT APPLICATION TRANSMITTAL

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Atty. Docket No. 8654-1090

9. ☒ Priority document(s) ☐ enclosed; ☒ to follow.
10. ☒ Statement of Small Entity Status, Pursuant to 37 C.F.R. §1.27
11. ☐ Microfiche Computer Program (Appendix).
12. ☐ Nucleotide and/or Amino Acid Sequence Submission.
☐ Computer Readable Copy.
☐ Paper Copy (identical to computer copy).
☐ Statement verifying identity of above copies.

13. Calculation of Fees:

FEES FOR	TOTAL CLAIMS	EXCESS CLAIMS	FEE	AMOUNT DUE
Basic Filing Fee (37 C.F.R. § 1.16(a))				\$710.00
Total Claims in Excess of 20 (37 C.F.R. § 1.16(c))	25	5	18.00	\$90.00
Independent Claims in Excess of 3 (37 C.F.R. § 1.16(b))	3	0	80.00	\$0.00
Multiple Dependent Claims (37 C.F.R. § 1.16(d))	Yes		270.00	\$270.00
Subtotal - Filing Fee Due				\$1,070.00
Reduction by 50%, if Small Entity (37 C.F.R. §§ 1.9, 1.27, 1.28)	0.50			\$0.00
TOTAL FILING FEE DUE				\$0.00
Assignment Recordation Fee (if applicable) (37 C.F.R. § 1.21(h))	0		40.00	\$0.00
GRAND TOTAL DUE				\$535.00

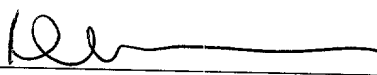
14. PAYMENT is:
☒ included in the amount of the **\$535.00** by our enclosed check.
☐ not included, but deferred under 37 C.F.R. § 1.53(f).

15. All correspondence for the attached application should be directed to:

Palmer & Dodge, LLP
 One Beacon Street
 Boston, MA 02108
 Telephone: (617) 573-0100
 Facsimile: (617) 227-4420

Respectfully submitted,

Date: 11/10/00

By: 
 Name:
 Registration No.: **34,380**

COMBINED DECLARATION AND POWER OF ATTORNEY

**(ORIGINAL, DESIGN, NATIONAL STAGE OF PCT, SUPPLEMENTAL, DIVISIONAL,
CONTINUATION, OR C-I-P)**

As a below named inventor, I hereby declare that:

TYPE OF DECLARATION

This declaration is for a national stage of PCT application.

INVENTORSHIP IDENTIFICATION

My residence, post office address and citizenship are as stated below, next to my name. I believe that I am an original, first and joint inventor of the subject matter that is claimed, and for which a patent is sought on the invention entitled:

“SELECTION SYSTEM”

SPECIFICATION IDENTIFICATION

The specification is attached hereto.

ACKNOWLEDGMENT OF REVIEW OF PAPERS AND DUTY OF CANDOR

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information, which is material to patentability as defined in 37, Code of Federal Regulations, Section 1.56.

PRIORITY CLAIM (35 U.S.C. Section 119(a)-(d))

I hereby claim foreign priority benefits under Title 35, United States Code, Section 119(a)-(d) of any foreign application(s) for patent or inventor's certificate or of any PCT international application(s) designating at least one country other than the United States of America listed below and have also identified below any foreign application(s) for patent or inventor's certificate or any PCT international application(s) designating at least one country other than the United States of America filed by me on the same subject matter having a filing date before that of the application(s) of which priority is claimed.

Such applications have been filed as follows.

**PRIOR PCT APPLICATION(S) FILED WITHIN 12 MONTHS
(6 MONTHS FOR DESIGN) PRIOR TO THIS APPLICATION
AND ANY PRIORITY CLAIMS UNDER 35 U.S.C. SECTION 119(a)-(d)**

INDICATE IF PCT	APPLICATION NUMBER	DATE OF FILING DAY, MONTH, YEAR	PRIORITY CLAIMED UNDER 35 U.S.C. SECTION 119
PCT	GB99/01526	13 May 1999	

**ALL FOREIGN APPLICATION(S), IF ANY, FILED MORE THAN 12 MONTHS
(6 MONTHS FOR DESIGN) PRIOR TO THIS U.S. APPLICATION**

<u>GB</u>	Patent App. <u>9810223.9</u>	filed <u>13 May 1998</u>
<u>GB</u>	Patent App. <u>9810228.8</u>	filed <u>13 May 1998</u>
___	Patent App. _____	filed _____

POWER OF ATTORNEY

I hereby appoint the following practitioner(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith.

APPOINTED PRACTITIONER(S)	REGISTRATION NUMBER(S)
Paula Campbell Evans	32,503
Kathleen M. Williams	34,380
Richard B. Smith	34,020
John M. Garvey	37,833
Mark J. Fitzgerald	45,928
Elizabeth Spar	45,123
Dianne M. Rees	45,281

**AUTHORIZATION OF ATTORNEY(S) TO ACCEPT AND
FOLLOW INSTRUCTIONS FROM REPRESENTATIVE**

The undersigned to this declaration and power of practitioner hereby authorizes the U.S. practitioner(s) named herein to accept and follow instructions from:

ATTORNEY
Palmer & Dodge LLP
One Beacon Street
Boston, MA 02108-3190

as to any actions to be taken in the Patent and Trademark Office regarding this application without direct communication between the U.S. practitioner(s) and the undersigned. In the event of a change in the person(s) from whom instructions may be taken, the U.S. practitioner(s) will be so notified by the undersigned.

SEND CORRESPONDENCE TO

ATTORNEY
Palmer & Dodge LLP
One Beacon Street
Boston, MA 02109-3190

DIRECT TELEPHONE CALLS TO:

ATTORNEY
617-573-0100

DECLARATION

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

SIGNATURE(S)

RIECHMANN, Lutz

Inventor's signature _____

Date _____

Residence Great Britain

Post Office Address 38 Beaumont Road, Cambridge CB1 8PY Great Britain

British

Country of Citizenship

KRISTENSEN, Peter

Inventor's signature _____

Denmark

Date _____

Country of Citizenship

Residence Denmark

Post Office Address Silkeborg WJJ J32 2th, DK-8000 Aarhus, DK

JESTIN, Jean-Luc

Inventor's signature _____

French

Date _____

Country of Citizenship

Residence France

Post Office Address 42 rue des Gravilliers, F-75003 Paris, France

WINTER, Gregory Paul

Inventor's signature _____

British

Date _____

Country of Citizenship

Residence Great Britain

Post Office Address Trinity College, D4 Neville's Court, Cambridge CB2 1TQ

Selection System

This is a continuation of International Application GB99/01526, with an international filing date of May 13, 1999, now abandoned. The invention relates to a selection system which permits the selection of polypeptides displayed in a phage display system.

Viruses have been used for the display of peptides and proteins [21, 26, 44]. In particular filamentous bacteriophage have been used for display of proteins and peptides by fusion of the genes encoding the proteins or peptides to the gene encoding a phage coat protein. As the fusion gene is encapsidated in the phage that is displaying the fusion protein, this provides a linkage of phenotype and genotype. Repertoires of proteins can be encoded by a population of phage, and the rare phage comprising proteins with predefined binding activities isolated by binding to solid phase. In this way synthetic human antibodies of predefined antigen-binding specificity have been selected from repertoires of antibody fragments assembled from different structural elements [10]. As the antibody needs to be folded to bind antigen, selection for binding also selects for folding. This principle has also been used for selection of folded peptides where binding is mediated by a discontinuous epitope [8, 11-13].

A problem present in phage display systems is the presence of high levels of background caused by the presence of phage not displaying desired polypeptides. For example, antibody repertoires are commonly encoded as fusion proteins with the p3 protein on phagemid vectors and are encapsidated by the use of helper phage. The helper phage coat protein competes with the fusion of antibody and coat protein (encoded on the phagemid), leading to phage with "monovalent" rather than multivalent display of folded antibody fragments. This can be useful in discriminating between the affinity and the avidity (with multivalent display) of the antibodies displayed on the phage. However the great majority of phages only display the helper phage coat protein which contributes to a "background" binding to antigen.¹ In this case it is desirable to select for phages that display folded antibodies, and to eliminate those that do not.

Moreover, all of the systems in current use rely on a binding activity in the polypeptide to be selected in order to perform the isolation of the desired display bodies from those

which do not encode polypeptides having a desired characteristic. This places a limitation on available display systems to the selection of folded polypeptides which possess a known binding activity. It would be desirable to have a means for selection of displayed proteins or polypeptides that is independent of the binding activity thereof.

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For example, there is considerable interest in building folded proteins *de novo*. Attempts have been made to design proteins *de novo* by assembly of predefined elements of secondary structure and also from random sequences (for review [5]). In some cases the designed proteins have been shown to retain elements of secondary
10 structure but lack the stable tertiary interactions characteristic of the folding of native proteins, suggesting the presence of molten globules (see [6] and references therein). More successful has been the creation of native-like proteins based on a pre-existing backbone [7, 8]. In these cases the binding activities of a *de novo* designed protein will be unknown. In this case it is desirable to select for phages displaying folded proteins,
15 and to eliminate those that do not.

Although attempts have been made to screen for folded proteins by their ability to survive degrading enzymes in bacteria [16-18], such methods do not allow for selection if bacterial growth or survival does not depend on the function of the folded protein.
20 Thus, these systems are only applicable to a small minority of polypeptides which one might wish to select according to the ability to fold.

It has previously been shown that the insertion of a peptide sequence between a proteolytically stable tag fused to the minor phage coat protein p3 and the p3 protein
25 itself, followed by proteolysis, provides a means to select for phages bearing peptide sequences that are susceptible to proteolysis [19, US Patent 5,780,279]. In these experiments, phage are bound to an affinity resin binding an N-terminal, proteolytically stable tag on the phage. If the bound phage are subjected to proteolysis and elution, only phage with cleavable sequences are eluted. This method is used to identify, among
30 a repertoire displayed on phage, amino acid sequences suitable as substrates for proteases. The sequences introduced are short and would not be capable of folding independently. Moreover, the system selects specifically for eluted rather than bound

phage; in other words, it is specifically configured to isolate cleaved rather than uncleaved phage.

Summary of the Invention

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The present invention exploits the application of peptide cleavage to eliminate unwanted viruses.

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According to a first aspect, therefore, the invention provides a method for the selection of a virus comprising the steps of:

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- (a) providing a virus encoding and displaying a fusion polypeptide, said fusion polypeptide comprising a heterologous polypeptide inserted into the sequence of a viral coat protein polypeptide, wherein said virus comprises a cleavable site located within a displayed polypeptide;
- (b) exposing the virus to a cleaving agent;
- (c) propagating the virus comprising intact fusion protein.

20

According to the present invention, virus may be selected by cleavage of non-resistant virions using a cleaving agent. As used herein, "virus" refers to an infective inoculum of virions, which may incorporate cleavage sites, optionally as part of heterologous polypeptides encoded by the viral genome. Thus, "virus" may refer to a plurality of virions, such that it may encode a repertoire of polypeptides; alternatively, as the context requires, it may be used to denote a single virion. The term "virus" includes any suitable virus which may incorporate a cleavage site, either naturally or through manipulation. A preferred virus for use in the present invention is bacteriophage, preferably filamentous bacteriophage.

25

30

The term "polypeptide" is used generally to denote molecules constructed of a plurality of amino acids, the amino acids being joined together covalently such as through peptide bonds. "Fusion" polypeptides are essentially polypeptides which are incorporated into viral coat proteins, such that a fusion is created between the viral coat protein and the polypeptide in question. The fusion may incorporate the polypeptide

into the viral coat protein, advantageously between domains thereof, or place it at one end thereof, to make a terminal fusion. The polypeptide is referred to as a "heterologous" polypeptide, to denote that it is heterologous to the viral coat protein into which it is inserted. It is possible, however, that it is derived from another polypeptide of said virus.

In one sense, polypeptide is used interchangeably with "protein" herein, in that no difference of structure or size is implied. Substantially any polypeptide may be selected for by the method of the present invention, including structural polypeptides, polypeptides having enzymatic activity and polypeptides having binding activity, including antibodies and antibody fragments. Cleavage sites may be present in the polypeptides, and may be naturally-occurring or may be engineered into the polypeptide or into a linker peptide attached thereto. "Polypeptide" may also refer to inserted polypeptides which are essentially non-folding polypeptides and serve to encode a cleavable site and insert this site into the coat protein of a virus. Inserted polypeptides may take the form of N- or C-terminal fusions, or may form part of the coat protein itself.

A "cleavable site" is a site capable of cleavage when exposed to a cleaving agent. In the present invention, the use of protease cleavage sites, capable of being cleaved with proteases, is preferred. Protease cleavage sites may be part of, or incorporated in, polypeptides according to the invention; alternatively, it may be independently engineered into the coat protein of the virus. A feature of the cleavable site is that it should either be absent from the virus other than at the site of its specific insertion according to the present invention, or otherwise inaccessible to cleavage, or present only in viral proteins which are not required after virion assembly to mediate infection.

In accordance with the invention, the cleavable site may be inserted into or present in any suitable position in the virus. Advantageously, however, it is inserted into or present in either the coat protein itself or the heterologous polypeptide which forms part of the fusion polypeptide.

In a preferred aspect of the present invention, more than one cleavable site may be used. For example, one site may be inserted or otherwise known to be present in the virus, whereas the presence of another site may be unknown or dependent on randomisation of the heterologous polypeptide sequence. In a particularly preferred
5 embodiment, the cleavable sites may comprise a protease cleavable site and a bond formed through sidechains on one or more amino acids, such as a disulphide bond. Disulphide bonds are cleavable by reducing agents, such as DTT or β -mercaptoethanol.

If a virus contains a disulphide bond, cleavage of a protease cleavable site located
10 between the two cysteine residues which form the bond through their sidechains will not lead to loss of viral infectivity since the disulphide bond is capable of retaining the covalent linkage of the viral polypeptides.

Thus, the invention further provides a method for identifying the presence of disulphide
15 bonds in polypeptides.

Conversely, in the event that the selection of disulphide-containing polypeptides is not desired, viruses are advantageously treated with a reducing agent before or after proteolysis, in order to eliminate background due to viruses which have been cleaved by
20 the protease but which have been held together by disulphides.

The fusion polypeptide may comprise one or more heterologous polypeptides. In the case of a terminal fusion, one such heterologous polypeptide may function as a protein tag, allowing phage which express the fusion polypeptide to be identified. The
25 cleavable site may, in such a case, be positioned in or near the tag, such that cleavage of the cleavable site releases the tag.

A tag is any suitable entity capable of binding to a ligand which may be used to isolate a virus by the method of the present invention. Accordingly, the tag is resistant to the
30 cleaving agent used in the method of the invention. Examples of tag/ligand pairs include barnase/barstar, avidin/biotin, antibody or antibody fragments and ligands, chelating groups and chelates, for example metals, and the like.

In all embodiments of the present invention, which are described in greater detail below, the uncleaved polypeptides are selected for; the cleaved material is discarded in the selection step.

- 5 Preferably, the virus according to the invention encode a repertoire of heterologous polypeptides. A repertoire is a collection of members, preferably polypeptides, which differ slightly from each other in a random or partially randomised manner. Preferably, a repertoire of polypeptides is a collection of variant polypeptides which preferably incorporate random or partially randomised mutations. As used herein, a repertoire
10 preferably consist of 10^4 members or more. A repertoire advantageously comprises a very large number of members, typically between 10^8 and 10^{11} , and potentially 10^{14} or higher.

- The heterologous polypeptide, or repertoire of such polypeptides, is advantageously
15 displayed on the surface of the virus which encodes it, by virtue of its being incorporated into a coat protein, or on the surface of cells infected by the virus. Where the virus is a bacteriophage, the protein may also be displayed on the surface of bacteria infected with the bacteriophage.

- 20 The cleavable site is advantageously located in or adjacent to the heterologous polypeptide, such that it can be protected by folding of the heterologous polypeptide and thus allow selection for heterologous polypeptides which are capable of correct folding. Alternatively, however, the cleavable site may be located distal to the heterologous polypeptide; in such embodiments, the cleavable site may serve to permit reduction of
25 background in phage display techniques. For example, introduction of the cleavable site into helper phage used with phagemid encoding a repertoire of polypeptides allows helper phage to be removed by cleavage prior to infection of host cells, thus dramatically reducing background due to "empty" phage. Advantageously, therefore, the cleavable site is incorporated into the virus coat protein.

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As referred to herein, a phagemid is a plasmid cloning vector which comprises viral replication sequences but is deficient in at least one viral function. This means that

whilst phagemid may be inserted into host cells by conventional nucleic acid transfer methods, and will exist in the host cells in an episomal state, they are unable to assemble into virions and thus complete a viral cycle of infection. Helper phage are used to supply the deficient viral functions and permit the phagemid to be packaged into virions. In accordance with the invention, phagemid may encode coat protein fusions with heterologous polypeptides which incorporate a cleavable site.

Helper phage provide the viral function lacking in phagemid in order to allow packaging of the phagemid into virions. According to the invention, helper phage may be modified in order to render them cleavable by a cleaving agent, for example a protease. In one aspect of the present invention, helper phage may incorporate a coat protein having a cleavable site which, when cleaved in the "rescued" progeny phage, will render the helper phage-derived coat protein unable to mediate infection.

As will be apparent from the forgoing, in the method according to the present invention the virus which are resistant to cleavage are selected. Advantageously, the resistant virions will be selected by infection of susceptible host cells, such as bacterial host cells. Cleaved virions are not infective. Alternatively, binding of virions to a ligand, for example via a tag, is dependent on protection of a cleavable site in the virus, such that viruses which are cleaved are not isolated by ligand/tag binding.

Detailed Description of the Invention

The present invention may be configured in a number of ways according to the intended procedure to which it is desired to apply the basic methodology. Selective cleavage of virions is used to reduce background in phage display techniques. By cleaving and removing phage which either do not contain a heterologous polypeptide, or express a heterologous polypeptide which is not capable of correct folding, the sensitivity of a phage display process can be increased substantially. As demonstrated in the experimental section below, using the methodology of the invention phage display may be used to select polypeptides which are not susceptible to selection by display techniques according to prior art methods.

In a first configuration, cleavage of a cleavable site in a virion coat protein may be exploited to reduce background attributable to phage displaying no heterologous polypeptides, or phage displaying heterologous polypeptides which are incapable of folding correctly. Cleavage of displayed polypeptides, in accordance with the present invention, results in the impairment of the viruses to achieve infection of host cells. Thus, by propagating viruses which have been exposed to a cleavage agent, it is possible to enrich the virus for virions which comprise displayed polypeptides which are resistant to cleavage. As used herein, "impair" means to reduce; it thus includes but partial and complete prevention of infection of host cells by affected virus.

The coat protein is selected as the site for cleavage on the grounds that it is available to cleaving agents in the host cells harbouring the virus or at the surface of the virus itself. Thus, in a preferred embodiment, virus preparations may be treated with a cleaving agent, in order to render virions having cleavable coat proteins unable to mediate infection of host cells. Alternatively, cells infected with the virus may be treated with a cleaving agent active within the cell, which will prevent packaging of virus comprising a cleavable coat protein.

According to the present invention, reference to selection may be interpreted as a reference to screening, since the same processes may be used to screen phage, as will be apparent to persons skilled in the art.

5 Cleavable sites may be naturally part of the coat protein, but preferably they are engineered therein. Preferred cleavable sites include protease cleavage sites, which may be found in polypeptides or engineered as an integral part of their sequence. Typically, protease cleavage sites may be defined in terms of amino acid sequences which are susceptible to cleavage by a protease. For example, the invention
10 encompasses the use of protease cleavage sites cleavable by one or more of the proteases trypsin (cleaves at Lys, Arg), chymotrypsin (Phe, Trp, Tyr, Leu), thermolysin (small aliphatic residues), subtilisin (small aliphatic residues), Glu-C (Glu), Factor Xa (Ile/Leu-Glu-Gly-Arg), Arg-C (Arg) and thrombin.

15 Protease cleavage sites may be incorporated into the coat protein of a virus by constructing a fusion between the coat protein and a further polypeptide, the further polypeptide containing the cleavage site. The further polypeptide should be inserted at a position in the viral coat protein such that it allows the assembly of a functional viral capsid and subsequent infection, but if cleaved will result in the impairment of
20 infectivity.

If the protease cleavage site incorporated in the coat protein remains uncleaved, therefore, the virus is capable of assembly into functional virions and retains the potential to infect host cells. If the protease cleavage site is cleaved, however, the
25 structure of the viral coat protein will be compromised and the viral will lose at least part of its potential to infect host cells.

In a preferred embodiment, the virus for use in the present invention is a bacteriophage, preferably filamentous bacteriophage. Filamentous bacteriophage is widely used in
30 phage display techniques for the selection of polypeptides from phage libraries encoding a large repertoire thereof. Conventionally, the repertoire of polypeptides is inserted in

the p3 protein of filamentous bacteriophage, but any other suitable site may be employed within the scope of the present invention.

In the case of the p3 protein of filamentous bacteriophage the protein consists of three domains. The N-terminal D1 is involved in binding to the tolA receptor, D2 in binding to the F-pilus (and mediating infection) and D3 in anchoring the protein to the phage particle. Peptides and proteins can be inserted at the domain boundaries without abolishing infectivity [21, 22], but the presence of all the domains is essential for phage infectivity [23]. The bacteriophage are resistant to proteolysis (allowing their use as "substrate" phage, [19]), but the introduction of polypeptides comprising protease cleavage sites into p3, for example at the junctions between domains leads to loss in infectivity of the phage upon proteolysis.

The protease cleavage sites may be incorporated into heterologous polypeptides. As described above, heterologous polypeptides may be encoded in the form of a repertoire in a phage library. As folded polypeptides or proteins are often resistant to proteolysis and unfolded proteins are sensitive, cleavage requires the polypeptide chain to bind and adapt to the specific stereochemistry of the protease active site, and therefore to be flexible, accessible and capable of local unfolding [14, 15]. The cloning of a polypeptide comprising protease cleavage sites at the domain junctions of p3, followed by proteolysis, provides a means of selection for phages bearing proteins that are resistant to proteolysis and are folded.

In the case of phage display repertoires (wherein the polypeptide to be selected is cloned at the N-terminus of p3) encoded on phagemid vectors, the use of helper phage comprising a polypeptide comprising protease cleavage sites at the domain boundaries, followed by proteolysis, provides a means of selection for phages that display the fusion protein by eliminating the helper phage after the "help" has been given.

The use of the protease-cleavable helper followed by protease cleavage selects for phages bearing the fusion protein (and for good display). As many phages in a repertoire do not display fusion proteins [26] and these contribute to non-specific

binding of the phage, this should also improve selection efficiencies. When using the techniques of the prior art, only between 0.1-1% of all phage particles in a phage library may comprise a gene 3 protein arising from the phagemid. Therefore the majority (99-99.9 %) of phage particles that have bound non-specifically to the solid support used in selection will comprise p3 from the helper phage (irrespective of the genome carried by the phage particle which most likely will be a phagemid DNA), these particles are rendered non-infective by proteolytic cleavage.

According to a third embodiment, the selection process may be used for identification of interacting protein elements. If two such elements linked by a polypeptide comprising protease cleavage sites are cloned between the D2 and D3 domains for display on phage, the only infectious phages after proteolysis are those in which the D2 and D3 domains are held together by non-covalent interactions between the interacting protein elements. The invention accordingly permits selection of a repertoire of polypeptides for its ability to interact with a selected polypeptide, or a second repertoire of polypeptides. Unlike the two-hybrid system, the invention relies on dissociation of non-interacting elements as distinct from the association of interacting elements for the selection step. Moreover, the invention permits the harnessing of the power of phage display to greatly increase the degree of selection.

The invention optionally comprises the use of conditions or agents, during cleavage of the cleavable site, which modulate the lability of the cleavage site in the presence of the cleaving agent. This approach may be used to increase cleavage, for example to select only for polypeptides which fold in such a manner as to closely shield the cleavable site from access by the cleaving agent, or to decrease cleavage, for example to select stable mutants from a repertoire of polypeptides which is ordinarily relatively labile under cleavage conditions.

For example, modulation of the lability of the cleavable site may be achieved by the use of agents which increase or decrease such lability. Thus, a protein denaturant may be included, at a suitable concentration, to destabilise a polypeptide and render it more labile. Alternatively, a ligand for a polypeptide may be included. The ligand may

stabilise the folded structure of the polypeptide, rendering it less sensitive to cleavage. Alternatively the ligand may destabilise the folded structure of the polypeptide, for example by favouring the adoption of an alternative configuration. This may render the polypeptide more accessible to the cleavage agent, and thus more labile.

5

In a further embodiment, the conditions of the cleavage process may be altered, such as by manipulating the pH or the temperature at which cleavage is conducted, to achieve similar effects. Thus, deviation of the pH from the optimum for the polypeptide comprising the cleavable site may cause the site to become more accessible to the cleaving agent. Similarly, raising (or lowering) the temperature of the conditions under which the polypeptide is cleaved may render the polypeptide more or less susceptible to cleavage.

In some instances, non-covalent interactions may be responsible for peptides retaining their structure and coat proteins remaining viable, even after successful cleavage of the cleavable site. The use of denaturants, temperature variation and other potentially destabilising techniques may also be used to decrease the likelihood of a cleaved polypeptide retaining its structure.

Proteolytic selection for protein folding may be applied in a number of areas, as it allows much larger numbers of proteins to be processed than with conventional screening. For example, it allows the isolation of mutant proteins with improved stability [1], for example from combinatorial libraries of mutants in which residues at several sites are varied simultaneously [39, 40] or from random mutants or by recombination [3, 4]. It also allows the isolation of novel proteins and architectures from large repertoires of sequences [16-18, 41]; and for improvement in folding stability over several rounds of mutation and increasingly stringent selection, much like the affinity maturation of antibodies.

A second configuration of the present invention concerns the use of tags to allow isolation of correctly folded heterologous polypeptides, exploiting the ability of correctly folded polypeptides to protect a cleavable site on to near to an associated tag,

The insertion of a polypeptide between the stable tag fused to the N-terminus of the viral coat protein and the coat protein itself, followed by cleavage, provides a means of selection for virus bearing proteins that are resistant to proteolysis and are folded. Thus only virions, whose inserted polypeptide is not degraded, will keep the tag fusion as part of their coat, and only these virions can therefore be captured by affinity purification using this tag. After elution the affinity captured phases from the ligand, these phages can be propagated and subjected to further rounds of the same selection procedure.

10 Alternatively, virions may be bound to an affinity matrix, comprising a ligand for the tag, prior to cleavage. The cleaving agent may subsequently be added, and only resistant phage will be retained on the matrix. These may then be eluted as required.

15 Suitable matrices include columns, beads and other surfaces to which a ligand for the tag is bound.

According to the present invention, reference to selection may be interpreted as a reference to screening, since the same processes may be used to screen phage, as will be apparent to persons skilled in the art.

20 Cleavable sites are substantially as described for the previous configuration of the present invention and are advantageously protease cleavable sites.

25 Cleavage requires the polypeptide chain to bind and adapt to the specific stereochemistry of the protease active site, and therefore to be flexible, accessible and capable of local unfolding [14, 15]. Folded polypeptides or proteins are often resistant to proteolysis, due to a relative inflexibility in their structure, whilst unfolded proteins remain sensitive.

30 As referred to above, the possible selection of polypeptides from a repertoire which, through variation or mutation, do not contain a recognition sequence for any particular protease used in this method, can be circumvented in two ways. For example, the use

of a cocktail of proteases with very distinct recognition sequences would ensure that all polypeptides should be cleavable, if not protected by their folded status. Alternatively, a phage repertoire of polypeptides to be selected could be partially denatured, such that the inserted polypeptide unfolds but the phage and the N-terminal tag remains intact.

- 5 Proteolytic digestion followed by affinity purification would remove all phages from the repertoire, which have escaped proteolysis due to the lack of protease recognition sequences in the polypeptide. Phages not bound by the resin, contain only phages, which contain the protease recognition sequence in the polypeptide displayed and which may or may not escape proteolysis under non-denaturing conditions. Thus these would
10 be subjected to proteolytic selection based on protection by the folding status of the polypeptide displayed.

- The selection process may also be used for the identification of interacting protein elements. Thus if two such elements linked by a polypeptide comprising protease
15 cleavage sites are cloned between the N-terminal, proteolytically stable tag for display on phage and the coat protein, the only phages after proteolysis, that can be captured via affinity binding to the tag, are those in which the tag and the p3 protein are held together by non-covalent interactions between the interacting protein elements.

- 20 The invention is further described in the following examples, for the purposes of illustration only.

Example 1. Resistance of filamentous phage to proteolysis.

- 25 Materials and Methods for Examples 1 – 6 are appended at the end of Example 6.

- Phage is incubated under a range of denaturing conditions *in vitro* and then restored to native conditions immediately before infection of bacteria. The incubation of phage in 10 M urea, or extremes of pH (as low as pH 2, and as high as pH 12) and temperature
30 (as high as 60°C) did not lead to a major loss of infectivity (Table 1). This indicates that the phage is either resistant to denaturing conditions or that if it does unfold it is

able to refold rapidly. However with GndHCl a 5 fold loss in phage infectivity is observed above 5 M and a further 5 fold loss at 8 M (Table 1).

Phage is then incubated under native conditions with a range of proteases (trypsin, Factor Xa, IgA protease, Asp-N, chymotrypsin, Arg-C, Glu-C, thrombin, thermolysin, subtilisin) with different specificities. There is no loss in infectivity except for subtilisin which has been reported to cleave the p3 protein [24]. If phage is incubated under denaturing conditions in the presence of proteases such as trypsin in 3.5 M urea (or > 47°C), infectivity is lost. This indicates that under denaturing conditions the unfolding of the phage coat proteins is sufficient to make sites available for proteolysis.

Example 2. Construction of phage with protease cleavage sites.

A sequence (PAGLSEGSTIEGRGAHE) comprising several proteolytic sites is inserted in the flexible glycine-rich region between the D2 and D3 domains of the phage p3. Incubation of the phage (fd-K108) under native conditions with trypsin, thermolysin or subtilisin now resulted in almost complete loss of infectivity (from 10^7 to <10 TU/ml) and incubation with Glu-C and chymotrypsin resulted in a major loss (from 10^7 to 10^4 TU/ml). This indicates that these proteases cleave the new linker. However incubation with Factor Xa, Arg-C or thrombin did not lead to a loss in infectivity, despite the presence of potential cleavage sites for these enzymes. Presumably the presence of the D2 and D3 domains may block access or cleavage for these enzymes in the case of the present polypeptide.

Example 3. Construction of protease cleavable helper phage and phagemid

Fusion of proteins to p3 should lead to multivalent display of the protein on the phage. However if the protein is fused to p3 encoded by a phagemid (such as pHEN1 [25]), and the bacteria harbouring the phagemid is rescued with a helper phage (such as VCSM13), the fusion protein has to compete for incorporation into the phage with the helper p3. This leads to so-called "monomeric" phage, in which usually less than one copy of the fusion protein is attached to each phage particle [26].

The use of "monomeric" phage might be expected to be advantageous for selection of high affinity interactions. Furthermore fusion proteins in "monomeric" phage should be more sensitive to proteolysis, as interactions between multimers of fusion protein are avoided. However a disadvantage is that the majority of infective phages do not display a protein; such phages binding non-specifically to solid phase are amplified during each round of phage growth.

Protease cleavable helper phage are therefore constructed, by introducing the protease cleavage sequence between the D2 and D3 domains to generate the helper phage KM13.

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Table 1. Stability of wild type fd-DOG under different conditions. The infectivity (TU/ml x 10¹⁰) was measured (see Materials and Methods) and has an estimated error of about \pm 6%.

Urea (60°C, 90 min)	Control	2 M	4 M	6 M	8 M	10 M	
	0.56	0.64	0.32	0.32	0.80	0.68	
GndHCl (37°C, 90 min)	Control	2 M	4 M	5 M	6 M	7 M	8 M
	0.72	0.60	0.70	0.16	0.13	0.16	0.03
pH (37°C, 30 min)	Control	pH 2.2	pH 4.0	pH 7.4	pH 10	pH 12	
	1.5	0.46	1.3	1.5	1.4	0.40	
Temperature (30 min)	Control	22°C	37°C	60°C			
	9.7	8.3	9.6	12.0			

KM13 is shown to rescue the phagemid pHEN1. Furthermore trypsin is shown to cleave a major fraction (about 50%) of p3 of the rescued phage as shown by Western blot and detected with an anti-D3 mAb (Fig. 1). However phage infectivity is hardly altered by the cleavage; it therefore appears that only a fraction of the p3 need be entire
 5 to mediate bacterial infection.

KM13 is also shown to rescue a pHEN1 phagemid encoding a single chain antibody fragment [27]. Here cleavage by trypsin resulted in a 50 fold loss in phage infectivity (data not shown), consistent with indications that only a small fraction of the phage
 10 express fusion protein when rescued with helper phage [26, 28].

A protease cleavable phagemid is also constructed. The phagemid can be rescued with KM13 or VCSM13. As expected, infectivity of this phagemid rescued with KM13 (but not VCSM13) is destroyed by trypsin. This phagemid vector is prone to deletions in the
 15 D2-D3 linker; by changing the codon usage in the linker regions on either site of the protease cleavable site, and shortening the length of these linker regions, a more stable vector is created (pK1; Fig. 2). In a second vector (pK2; Fig. 2), the sequence of the polylinker is arranged so as to place D3 out of frame to render religations within the polylinker non-infectious.

20
Example 4. Construction of a phage antibody library using protease cleavable helper phage.

Bacteria are electroporated with phagemid DNA encoding a repertoire of scFv
 25 fragments fused to the N-terminus of p3 and grown in liquid culture (2xTY containing antibiotic to select for bacteria containing phagemid and glucose to suppress expression of gene 3). In the mid log phase of bacterial growth ($OD_{600} = 0.5$) the helper phage KM13 is added to the bacteria to give a ratio of helper phage to bacteria of 20:1. The bacteria is incubated at 37°C without shaking for 45 min then with shaking for 45 min.
 30 The bacteria are harvested by centrifugation and resuspended in fresh medium containing 50 µg/ml kanamycin and antibiotic, without glucose, to select for presence of phagemid DNA. The culture is grown overnight at 30°C with shaking.

Bacteria are removed from the phage containing supernatant by centrifugation. Phage is precipitated from the supernatant by adding 1/5 the volume of 20% PEG/ 2.5 M NaCl. After 1-2 hours at 4°C the precipitated phage is collected by centrifugation. The phage is resuspended in PBS (a second PEG precipitation is optional) and can be used in selection.

The library of phages is allowed to bind to antigen (immobilised on solid support such as an immunotube or in solution to tagged (i.e. biotinylated) antigen which can be immobilised after affinity binding of phage antibodies). Unbound phage is removed by extensive washing (the stringency of washing can be varied with respect to time and detergents added).

Phage libraries comprising a cleavable tag, such as the c-myc tag inserted between the antibody and gene 3, can be eluted by addition of trypsin in solution at a concentration of 0.1 to 1 mg/ml. (Phage libraries without a cleavable sequence between the antibody and gene 3 can be eluted by adding 100 mM Triethylamine. In this case the solution is neutralised by adding 1 M Tris-HCl pH 7.4. and after 10 min, trypsin added to a final concentration of 0.1 to 1 mg/ml.) Trypsin also cleaves the copies of gene 3 from the helper phage, while leaving gene 3 from the phagemid intact. Thus only phage that had carried, or still carry, an antibody fusion will be infective.

Phage is used to infect bacteria in mid log phase of growth ($OD_{600} = 0.5$), and the bacteria is plated on agar plates containing antibiotic selecting for phagemid DNA. Individual clones were picked and phage prepared as above. The resulting phage is used in ELISA to identify phage antibodies binding specifically to the antigen of interest.

Example 5. Selection for folding using barnase as a model.

Barnase is a small RNase of 110 amino acid residues whose folding has been extensively studied (for review [2]). Barnase contains multiple sites for trypsin cleavage, although the folded protein is resistant to cleavage (data not shown). Phage

with barnase cloned between D2 and D3 should therefore be resistant to protease cleavage and capable of selection.

As barnase is toxic to *Escherichia coli*, a mutant A (His102->Ala) is cloned which is catalytically inactive but stable [29, 30] into the phagemid pK2. A mutant B (His102->Ala,Leu14->Ala) is also cloned, with lower stability; Leu14 is buried in the hydrophobic core and its mutation creates a large cavity in the core affecting the packing of different structural elements [31]. The phages (rescued with KM13) bind to the inhibitor barstar by ELISA (Fig. 3), and therefore display the mutant barnase in a folded form.

The phages are then incubated with trypsin at a range of temperatures (Fig. 4). After incubation at 10°C, there is a decrease in phage infectivity of 5 to 10 fold for both mutants, suggesting that (as above with the display of scFv fragment) only a small fraction of the phages display the fusion protein. There is no further loss in infectivity on cleavage until 30°C (for mutant B) or 37°C (for mutant A). In both cases the major transition is at least 10°C below that expected for the reversible thermal unfolding of the mutants.

Phages A and B are mixed in different ratios and incubated with trypsin at 20°C, where both mutants are stable to cleavage, or at 37°C where only A is stable. After "proteolytic selection" the phages are plated and analysed by PCR, which is followed by restriction digest to distinguish the mutants. As shown in the Table 2, mutant A is enriched by a factor of 1.6×10^4 after a single round and by 1.3×10^6 after two rounds of proteolytic selection at 37°C. No enrichment can be detected at 20°C.

Example 6. Selection for folding using villin as a model.

The 35 amino acid subdomain of the headpiece domain of the f-actin-bundling protein villin [32] is much smaller than barnase. It nevertheless forms a stable fold at room temperature and is resistant to proteolysis; furthermore its stability does not rely on disulphide bonds or binding ligands [33]. The villin subdomain (which contains several

potential trypsin cleavage sites) is cloned between the D2 and D3 domains of the phage, and incubated with trypsin at different temperatures. The profile for loss of infectivity is not as sharp as with barnase, with the major transition below 35°C, considerably below the thermal unfolding of villin (70°C) [32, 33]. The phage displaying villin are mixed
 5 with phage, which were produced using the phagemid pK1 and the helperphage KM13, and incubated with trypsin. After a single round of proteolytic selection, the villin fusion phage are enriched 8.7×10^3 fold (Table 3).

In summary, the results from Examples 1 and 2 show that the infectivity of the phage is
 10 relatively resistant to temperature, pH, urea and GndHCl, and to several proteases, but if a flexible linker comprising a protease cleavage site is inserted between domains D2 and D3 of the phage coat protein p3, the phage becomes sensitive to cleavage. By contrast, as shown in Examples 5 and 6, if the protease cleavage sites comprise a folded protein domain such as barnase or villin, the phage is resistant to cleavage. This allows
 15 proteolytic selection for protein folding, with enrichment factors of greater than 10^4 fold for a single round of selection. Selection is evident for both for barnase, an average sized [34] domain of 110 amino acids and for villin, a small domain of 35 amino acids.

Discrimination between structures of different stabilities can be accomplished by
 20 increasing the stringency of proteolytic selection. Thus with increase in temperature, both barnase and villin became susceptible to cleavage, reflecting protein unfolding. However the main impact of protease cleavage is at a temperature lower than the unfolding transition as measured by circular dichroism [38]. This may reflect the fact that the unfolding transition is a fully reversible process, whereas the cleavage by
 25 proteases (of unfolded structure) is a kinetic and irreversible process, pulling over the equilibrium from folded to unfolded (and cleaved) structure. This is consistent with the CD unfolding transition seen with villin [33], where at temperatures as low as 35°C there is evidence of unfolding, the same point at which villin starts to become susceptible to protease attack.

Table 2. Selection of Barnase mutants.

Mixtures of barnase mutants (A+B) in ratios from 1:1 to 1:10⁸ were selected by proteolysis at 37 °C, 24 (or 36 in round 2) phage clones analysed and numbers of each

- 5 mutant noted above. ^a Selection at 20 °C where both mutants are expected to be stable, ^b before selection.

Phage A: Phage B		1:1 ^a	1:10 ²	1:10 ⁴	1:10 ⁶	1:10 ⁸
Round 1	Phage A	16 (14 ^b)	24	20	0	nd
	Phage B	8 (10 ^b)	0	4	24	nd
	Enrichment	-	-	1.6x10 ⁴	-	nd
Round 2	Phage A	nd	nd	nd	24	0
	Phage B	nd	nd	nd	12	36
	Enrichment	nd	nd	nd	1.3x10 ⁶	-

Table 3 Selection of villin.

Mixtures of villin-phage and pK1 rescued with KM13 in ratios from 1:1 to 1:10⁶ were selected by proteolysis at 10°C, 24 phage clones analysed and number of each noted

5 above. ^a before selection.

villin-phage : pK1

	1:1	1:10 ²	1:10 ⁴	1:10 ⁶
pK1	0 (16 ^a)	0	7	24
Villin	24 (8 ^a)	24	17	0
Enrichment	-	-	8.7x10 ³	-

10

15

20

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Table 4. Primer sequences

pklinker	5'GGCACCCCTCAGAACGGTACCCACCCCTCAGAGGCCGGCTGGGCCGC CACCCCTCAGAG 3'
polyXafor	5'GGTGGCGGCCAGCCGGCCTTTCTGAGGGGTGCGACTATAGAAGGAC GAGGGCCCAGCGAAGGAGGTGGGGTACCCCTTCTGAGGGTGG 3'
polyXaback	5'CCACCCTCAGAAGGGGTACCCACCTCCTTCGCTGGGCCCTCGTC CTTCTATAGTCGACCCCTCAGAAAGGCCGGCTGGGCCGCCACC 3'
fdPCRBack	5'GCGATGGTTGTTGTCATTGTCGGC 3'
LIBSEQfor	5'AAAAGAAACGCAAAGACACCACGG 3'
LIBSEQback	5'CCTCCTGAGTACGGTGATACACC 3'
LSPAfor	5'GTAAATTCAGAGACTGCGCTTCC 3'
LSPAback	5'ATTTTCGGTCATAGCCCCCTTATTAG 3'
Flagprimer	5'CAAACGGGCGGCCGAGACTACAAGGATGACGACGACAAGGAACT GTTGAAAGTTGTTTAGCAA 3'
RECGLYfor	5'CCCCTCAGAAAGGCCGGCTGGGCCGCCGCCAGCATTGACAGGAGGT TCAGG 3'
RECGLYback	5'GAAGGAGGTGGGGTACCCGGTTCCGAGGGTGGTTCCGGTTCGGTG AT TTTG 3'
delCKpn	5'CCCTCGGAACCGGTACCCAGCTGCTTCGTGGGCCC 3'
Barnasefor	5'CTGGCGGCGGCCAGCCGGCCCTGCACAGGTTATCAACACGTTTGA C 3'
BarnaseH102Aba ck	5'CTCGGAACCGGTACCTCTGATTTTGTAAAGGTCTGATAAGCG 3'
villinfor	5'GGCGGCCAGCCGGCCTTTCTCTCTCTGACGAGGACTTCAAGGC 3'
villinback	5'CCTCGGAACCGGTACCGAAGAGTCCTTTCTCCTTCTTGAGG 3'

Materials & Methods (Examples 1 – 6)

Materials

5 All restriction enzymes, T4 ligase are obtained from New England Biolabs. Taq DNA polymerase is obtained from HT Biotechnology. Pfu DNA polymerase is obtained from Stratagene. Ultrapure dNTP from Pharmacia. Proteases and the protease inhibitor Pefabloc are obtained from Boehringer Mannheim, except chymotrypsin and trypsin TPCK-treated which are obtained from Sigma. All other chemical are likewise obtained
10 from Sigma.

Phage preparation

Escherichia coli TG1 [42] is used for cloning and propagation of phage. TG1
15 harbouring fd-DOG [43] or derivatives is grown overnight in 2xTY containing 15 µg/ml tetracycline. Phagemids are rescued using KM13 or VCSM13 as described [27]. Phage particles are prepared by two PEG precipitations [44].

Vector construction

20 The phage vector fd-DOG [43] is used as parent vector for construction of the protease cleavable fd-K108. Unique restriction sites (SfiI, KpnI) are introduced into the glycine rich spacer region between D2 and D3 using the Sculptor *in vitro* mutagenesis system (Amersham) and the oligonucleotide pklinker (Table 4). Further restriction sites (ApaI,
25 SalI) and sequence encoding a protease cleavage site are cloned between the SfiI and KpnI sites using the oligonucleotides polyXafor and polyXaback to create the vector fd-K108.

30 The protease cleavable helper phage KM13 is prepared from fd-K108 by transplanting into the helper phage VCSM13 a BamHI-ClaI fragment generated by PCR and primers fdPCRBack and LIBSEQfor.

A protease cleavable phagemid vector is derived from fd-K108 much as above except using pCANTAB 3 (Pharmacia). A FLAG-tag is introduced at the N-terminus of D1 by cloning of a NotI-SfiI fragment generated by PCR and primers Flagprimer and LSPAback. To circumvent deletions due to repeated sequence in the D2-D3 linker, the
 5 codon usage of the polylinker region is changed in two steps (a) using a Bam-SfiI fragment generated by PCR and primers RECGLYfor and LIBSEQfor, screening recombinants by PCR and the primers LSPAfor and LSPAback, (b) using a KpnI-ClaI fragment generated by PCR and the primers RECGLYback and LIBSEQback, screening recombinants using LSPAfor and LSPAback. The resulting vector is pK1. The entire
 10 p3 gene is sequenced using PCR cycle sequencing with fluorescent dideoxy chain terminators (Applied Biosystems) [45]. The "out of frame" vector pK2 is derived from pK1 by site direct mutagenesis using the oligo delCKpn and the Sculptor Amersham kit. The precise sequences of pK1 and pK2 are set forth in Kristensen *et al.*, (1998) Folding & Design 3: 321-328.

15 Cloning of Barnase and Villin

The vectors encoding the single barnase mutants, His102->Ala and Leu14->Ala [29, 46] are used as templates for PCR amplification with primers Barnasefor and
 20 BarnaseH102Aback and Pfu polymerase. The PCR products (encoding the single mutant His102->Ala, and the double mutant His102->Ala, Leu14->Ala) are digested using the restriction enzymes SfiI and KpnI, and ligated into vector pK2 to give the phagemids pK2BA and pK2BB respectively and the barnase genes sequenced using PCR cycle sequencing.

25 The 35 amino acid thermostable fragment of the headpiece of the f-actin binding protein villin [33] is amplified from chicken bursa cDNA using PCR primers villinfor and villinback with Pfu polymerase. The PCR products are cloned as above to give the phagemid pK2V.

Resistance of phages to denaturants, pH and proteases

For resistance to denaturants, 10 M urea in PBS (25 mM NaH₂PO₄, 125 mM NaCl pH 7.0) or 8 M GndHCl (Guanidine hydrochloride) and 50 mM Tris-HCl pH 7.4, 1 mM CaCl₂ (buffer A) is added to 10 μ l phage stocks (10^8 - 10^{10} TU) to give a volume of 1 ml and the conditions specified in Table 1. The phage are incubated for 1-2 hrs, then 100 μ l aliquot added to 1 ml TG1 (OD₆₀₀ ~ 0.5) and serial dilutions plated on TYE plates with 15 μ g/ml tetracycline. For resistance of phage to extremes of pH (2-12), Tris glycine or Tris HCl buffers (0.1 M glycine or 0.1 M Tris respectively) are added to 10 μ l phage stocks, and to neutralise each 100 μ l aliquot we added 50 μ l 1 M Tris-HCl pH 7.4 before infection. For resistance to temperature, buffer A is added to 10 μ l phage stocks to give a volume of 1 ml and incubated at a given temperature (20 - 60 C) for 1 hr. 100 μ l aliquots are added to TG1 and plated as above. For resistance to proteases, 100 mM NaCl, 50 mM Tris-HCl, 1 mM CaCl₂ pH 7.4 (Factor Xa 100 ng/ml or trypsin, chymotrypsin, thrombin, thermolysin and subtilisin all 100 μ g/ml) or 50 mM Tris-HCl, 1 mM EDTA pH 7.4 (IgA Protease 10 ng/ml) or 50 mM NH₄CO₃ pH 8.0 (Arg-C 100 μ g/ml, Glu-C 100 μ g/ml) or 25 mM NaH₂PO₄, 125 mM NaCl pH 7.0 (AspN 40 ng/ml) is added to 10 μ l phage stocks (fd-DOG and fd-K108) to give a volume of 100 μ l and a final concentration of protease as indicated. Digestions are incubated for 15 min at room temperature, samples (100 μ l) are then infected into TG1 as above.

For resistance to proteases in the presence of denaturants samples are prepared as above for urea and temperature denaturation. To 90 μ l aliquots 10 μ l trypsin (1 mg/ml) is added, after 5 min at room temperature 4 μ l Pefabloc (100 mM) is added and the samples are infected into TG1 as above.

Western blot

Phages (pHEN1 rescued using KM13 and pK1 rescued using VCSM13) are subjected to SDS-PAGE [47] before or after cleavage by trypsin (50 ng/ml). After semidry transfer

to PVDF membranes the filter is process essentially as described [27]. The primary antibody, monoclonal anti-gIII (MoBiTec), is added in a 1:5000 dilution followed by anti-mouse HRP-conjugated antibody (Sigma) in a dilution of 1:50000. Finally the filter is developed using the luminol based Chemiluminescence Western Blotting kit (Boehringer Mannheim).

ELISA

Phage displaying barnase mutants are analysis for binding to the RNase inhibitor barstar as described [44]. 10 pmol biotinylated barstar is mixed with approximately 10^{10} phage displaying barnase mutant A or barnase mutant B or villin or buffer A. Phage binding barstar is captured on Streptavidin coated plates (Boehringer Mannheim) and developed using HRP conjugated anti-M13 antibody (Pharmacia) and 2,2'-Azino-Bis(3-Ethylbenzthiazoline-6-sulfonic acid) (Sigma). Absorbance readings are taken at 405 nm.

Temperature denaturation

At each temperature approximately 10^{10} phage displaying the barnase mutants or villin (ampicillin resistant) is mixed with a cleavable control fd-K108 (tetracycline resistant), and a non-cleavable control phagemid, a chloramphenicol resistant derivative of pHEN1, rescued with KM13 in a total volume of 90 μ l of buffer A. After equilibration for 20-30 min at the temperature indicated, 10 μ l trypsin (5 μ g/ml) is added and the incubation continued for 2 min. Trypsin is neutralised by adding 4 μ l 100 mM Pefabloc. Infection and serial dilution is performed in TG-1 as above and aliquots are plated on TYE plates containing 100 μ g/ml ampicillin + 1% glucose, 30 μ g/ml chloramphenicol + 1 % glucose or 15 μ g/ml tetracycline.

Selection experiments

10 μ l of serial dilutions of the barnase mutant phage A is mixed with 10 μ l of the non-diluted barnase mutant phage B in 70 μ l buffer A. After 30 min incubation at 20°C or 37°C 10 μ l trypsin (5 μ g/ml) is added. Following 2 min. of digestion 4 μ l Pefabloc

(100 mM) is added. The phage are infected into TG1 as above. A second round of selection are performed by scraping bacteria in 3 ml 2xTY, 50 μ l inoculated into 50 ml 2xTY/Amp/Glu and the phagemid rescued and phage prepared as above. Clones are analysed by PCR using the primers LSPAfor and LSPAback followed by restriction digestion using DdeI.

Selections between pK2V and pK1 phage particles are performed as above, except the selection is performed at 10°C. Clones are analysed by PCR using the primers LSPAfor and LSPAback.

Example 7. Use of protease-cleavable helper phage for selection of signal sequences.

Translocation of proteins is directed by signal peptides [48]. These are known to share common features such as a positively charged amino-terminal region, a hydrophobic sequence and a carboxy-terminal region including the signal peptidase cleavage site. Signal peptides are involved in "an array of protein-protein and protein-lipid interactions" [49]. The signal sequence may in addition interfere with the protein folding pathway. They are also involved in translational regulation, mainly through the downstream box.

Enzymes such as DNA polymerase are very poorly displayed on phage particles, making their selection almost impossible. In order to improve the capabilities of phage display to select polymerases, therefore, improved signal sequences are designed and selected for using a phage display technique in which "empty" phage background is eliminated by digestion of helper phage.

Design of a signal sequence for optimal polymerase display on phage is not easily achieved. A selection strategy is therefore devised to isolate signal sequences from a library where mutations are introduced at selected sites. Although the signal sequence is not present on phage, its sequence is easily retrieved by sequencing the phagemid located within the phage particle.

Two libraries are generated from pelB and g3 leader sequences, making use of the following oligonucleotide primers for PCR amplification:

1: TACGCCAAGCTTGCATGC;

5 2: CTGCACCTGGGCCATGG;

3: GATTACGCCAAGCTTTG

4: GATTACGCCAAGCTTGCATGCANNDCTNTDTCAAGGAGACAGTCATAATG
ARRNNBCTATTGSYAAYSYASYAGBNTTGTTATTACTCSYANYCVNNCY
GDCCAATGGCCCAGGTGCAGCTG;

10 5: GATTACGCCAAGCTTTGNNNNCTTTTTWWGGAGATTTTCAACRTGARAAR
ATTATTATTCSYAATTSTYTTTAGTTSYTSYTTTCTWTGYGGYCCAGCCGGCCAT
GGCCCAGGTGCA.

6: CTTTATGCTTCCGGCTCG.

7: CGGCCCCATTCAGATCC.

15

The restriction sites HindIII and NcoI are noted in italics.

Library I deriving from the pelB leader and library II deriving from the g3 leader are prepared by PCR amplification of 4 (pelB) amplified with 1 and 2 and of 5 (g3) amplified with 3 and 2 respectively. Each PCR product is digested with HindIII and NcoI and purified with a gel extraction kit (Qiaquick, Qiagen). 0.2 µg of each resulting insert is mixed for ligation with about 2 µg of pHEN1-Stoffel vector (Fig. 6) previously digested with HindIII and NcoI and dephosphorylated with alkaline phosphatase (Boehringer Mannheim). The ligation mixture is purified by phenol-chloroform extraction and ethanol precipitation prior to electroporation into freshly prepared *E. coli* TG1.

Randomisation of 32 and 20 bases for the pelB and g3 leaders respectively is carried out: (i) near and within the Epsilon sequence just upstream the Shine-Delgarno sequence (ii) downstream the Shine-Delgarno sequence near and within the Downstream box [50] (iii) within the leader peptide at the N-terminal region containing the positively charged

amino acid residues, in the hydrophobic region [51] and in the C-terminal region close to the highly conserved peptidase cleavage site [52].

Phage is produced as described previously [25] except that the helper phage KM13 (Example 3) is used instead of VCSM13 and that 0.1 mM IPTG is added when specified to the overnight culture at 30°C. Selections for resistance to trypsin and for binding [44] is done as described earlier, except that 11 µg of anti-Taq antibody (Taqstart, Clontech) is coated overnight on immunotubes (Nunc). PCR screening is done with primers 6 and 7 using single *E.coli* TG1 colonies containing the phagemid as template following a previously described protocol; after gel electrophoresis on a 2% agarose gel. The size of the amplified fragment is used as a criterion to establish whether deletions within the polymerase gene have occurred.

The calculated diversity of the libraries computed from the degeneracy of the synthesised oligonucleotides is about $3.7 \times 10^{13} = 4^9 \times 3^7 \times 2^{16}$ and $1.7 \times 10^7 = 4^4 \times 2^{16}$ for the pelB and g3 leaders respectively. After transformation of *E.coli* with the phagemid libraries, the library size measured as the number of ampicillin-resistant colonies is found to be 1.3×10^7 and 9.6×10^6 for the pelB and g3 leaders respectively.

The selection for display of the polymerase is done by cleaving specifically the helper phage p3 copies with the protease trypsin so as to render non-infective all phage particles that are not expressing any p3-polymerase fusion protein.

Both libraries are mixed and the selection rounds are carried out in two conditions, with or without 0.1 mM IPTG in the culture medium. With IPTG, deletions of the polymerase gene or of part of it are noticed after round III (4 out of 28 clones) as shown by a PCR screening (see above); after round IV, these clones represent most of the population (28 out of 30). Without IPTG, these clones represent a significant part of the selected ones after round VI (3 out of 12). The selection is therefore changed from round five on by introducing in addition to selection for protease resistance, a selection for binding to an anti-Taq antibody. After the selection rounds VII and VIII, (3 out of

13) and (0 out of 19) clones respectively correspond to deleted p3-polymerase fusion proteins.

For characterisation of the leaders, the HindIII-NcoI fragments are subcloned after PCR
 5 amplification of individual *E.coli* colonies. The resulting phagemids are noted pHEN1-
 lx/Stoffel subcloning with $x=7,9,10$ and 12. The HindIII-NcoI and the NcoI-NotI
 fragment corresponding to the Stoffel fragment is sequenced on both strands using a
 373A DNA sequencer (Applied Biosystems).

10 For ELISA, an anti-Taq antibody (Taqstart, Clontech) is used for coating the ELISA
 plate and an anti-M13-horseradish peroxidase fusion protein (Pharmacia Biotech) is
 used for detection in a standard protocol [44].

Expression of polymerase in the supernatant is made by infection of *E.coli* HB2151
 15 with selected phagemids [25, 27] except that the IPTG concentration is 0.1 mM instead
 of 1 mM. About 10 μ l of supernatant is loaded on a polyacrylamide gel for
 electrophoresis (Novex); the gel is blotted onto nitrocellulose (Protran, Schleicher and
 Schuell) and an anti-Taq polymerase antibody (Taqstart, Clontech), and a goat anti-
 mouse IgG-horseradish peroxidase (Sigma) prior to detection on autoradiography films
 20 by chemiluminescence (ECL reagents, Amersham).

Four individual clones 7, 9, 10 and 12 from round VII, that were screened for protease
 resistance among 12 clones, are further characterised. To ensure that only mutations
 within the signal sequence are considered, and not mutations somewhere else within the
 25 phagemid that may have occurred during amplification at the various rounds, the signal
 sequences are subcloned into the original vector. The results shown in Table 5 indicate
 that optimal polymerase display for the selected clone 10 is about 50 fold higher than
 for the original sequence. This result is confirmed independently within experimental
 errors by an ELISA using anti-Taq antibody and anti-M13-HRP: 10^9 phage particles of
 30 the pelB leader phagemid pHEN1-Stoffel (signal to noise ratio: 1.46) give an identical
 signal as 10^7 phage particles of clone 10 (signal to noise ratio: 1.47).

The expression of Stoffel fragment in *E.coli* HB2151 is also studied for the various leaders (see Table 6). The concentration of Stoffel fragment in the culture supernatant is estimated by comparing the spot intensities for known amounts of polymerase and found to be about 0.1 mg/l for the pelB leader. A 3-fold increase in expression is observed for the leaders l7 and l10, whereas an about 3-fold decrease is noted for leader l9.

Table 5. Number of phage-polymerases per phage particle with leader pelB, as a function of temperature and IPTG concentrations.

The phage titer is measured as the number of infective phage particles and the phage-polymerase titer as the number of infective phage particles after treatment with trypsin. The number of phage-polymerases per phage particle is the ratio of the titers. As the phage particles were rescued with a helper phage, phage displays either a p3-polymerase fusion protein and a few p3 copies containing a trypsin-cleavage site or only these p3 copies.

Temperature	25°C	30°C	37°C
0 mM IPTG	1.7×10^{-3}	9.1×10^{-4}	1.2×10^{-3}
0.1 mM IPTG	9.1×10^{-3} *	8.3×10^{-3}	2.4×10^{-3} *
1 mM IPTG	1.5×10^{-2} *	5.5×10^{-3} *	1.2×10^{-3} *

* in these conditions, the phage titer drops below 10^{10} /ml of culture medium.

Table 6. Phage characterisation for leader pelB or selected leaders from round VII for optimal polymerase display (same legend as for Table 5; culture in 2xTY at $T=30^{\circ}\text{C}$ without IPTG).

5	Leader	Titer	Number of phage-polymerases per phage particle
		$\times 10^{11}$	
	pelB	1.2	9.1×10^{-4}
	17	2.0	2.5×10^{-2}
10	19	0.5	8.3×10^{-3}
	110	0.7	4.3×10^{-2}
	112	1.6	1.4×10^{-2}

Table 7. Randomised and selected sequences.

The randomised DNA sequence is given from 5' to 3'; above and below it, the bases that differ from the given sequence in the signal sequences pelB, 17, 19, 110 and 112 are indicated. The Shine-Delgarno sequence, the start codon and the last codon of the signal sequence, GCC, have been underlined. The HindIII and the NcoI restriction sites are in italics. The corresponding amino acid sequences are given below. Library I is initially designed from the pelB leader and library II from the g3 leader.

10 III-A. From library I

pelB AATT A T AATAC
 5' AAGCTTGCATGCANNDCTNT DTCAAAGGAGACAGTCATAATGARRNNB CT
 17 GCAT C G AGACG
 15 110 CGGG G T GAGGG
 112 CCAG C T GCCGG

pelB CCT CGGC GCCGCT GA GCGGC CAG C G
 20 ATTGSYAAYRSYASYASYAGBNTTGTTATTACTC SYANY CVNNCYGDCCATGG
CC 3'
 17 GC TGGT CT GT GA CC CC GGT C T
 110 GC TGCT GT GC GG CC AT GCG C G
 112 GT TAGC GC GT GG CT GC CCC C A

25
 pelB M K Y L L P T A A A G L L L L A A Q P A M A
 17 K T A M V L V G P P G P S
 110 R G A M L V A G P I A P A
 112 R R V I A A V G L A P P¹ T

III-B. From library II

g3 leader GAGC TT G A A

5' AAGCTTTGNNNNCTTTTTTWGGGAGATTTTCAACRTGARAARATTATTAT

5 19 GGGC TA A G G

GC CC GT CC A C C

TCSYAATTSYTTTGTTSYTSYTTTCTWTGYGGYCCAGCCGGCCATGG CC3'

19 CT CC GT GC A T T

10

g3 leader M K K L L F A I P L V V P F Y A A Q P A M A

19 R R L P V A Y V V

Example 8. Selection of a catalytic activity using protease-cleavable helper phage.

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A strategy for the selection of catalysts by phage display is based on selection of the reaction product of a catalytic reaction, and the use of proximity effects to select the catalyst. In this strategy, a tagged substrate is crosslinked to the phage in the proximity of the displayed enzyme; the phage is thereby attached to a solid-phase and released by an intramolecular cleavage reaction catalysed by the displayed enzyme [53].

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A similar approach has been applied to the selection of active DNA polymerase variants. The approach involves two chemically independent reactions, the catalytic reaction leading to a product (in this case distinguished by incorporation of a biotin tag) and a chemical crosslinking reaction by which the substrate (and product) are linked to the phage. Selection of the phage by streptavidin beads therefore selects for phages which are chemically attached to tagged product; these reactions are more likely to be coupled on the same phage as reactions in cis are favoured over reactions in trans by proximity.

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Maleimides are used in a chemical cross-linking reaction. These are known to react with thiols and in alkaline solutions with amino groups, and are therefore capable of

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fusion protein are infective; from the loss in titre after proteolysis we estimate that only one phage particle in a thousand had incorporated the fusion protein. The selection process, relying on tagging by polymerase in cis, would be compromised by such a great excess of phages lacking the polymerase but available for tagging in trans.

- 5 Selected phages are therefore treated with trypsin to destroy the infectivity of those lacking the displayed polymerase.

The phage displaying the Stoffel fragment are incubated with primer 13 [TTT CGC AAG ATG TGG CGT] comprising a 5' maleimidyl group and a 3' biotinylated
 10 nucleotide. After incubation the phage are captured on streptavidin-coated beads, with a yield of about 1 - 5 % of infectious phage. This shows that primer can be chemically cross-linked to the phage, presumably via p8 protein as shown for the N-biotinoyl-N'-(6-maleimidohexanoyl) hydrazide. The phage are then incubated with primer 1b [GCGAAGATGTGG] comprising a 5' maleimidyl group in the presence of biotin-dUTP 2
 15 and template 3 [AAA TAC AAC AAT AAA ACG CCA CAT CTT GCG]. Capture of the phage is dependent on presence of 1b, 2 and 3 (Table 8), but also on the inclusion of trypsin, which cleaves the helper phage to reduce non-specific phage isolation.

Table 8. Selection of catalytically active phage-Stoffel particles.

	ϕ_i [a]	ϕ_f [b]	Yield	Conditions [c]
	in tu	in tu	in %	
5	8.4×10^5	2.0×10^4	2.4	
	3.6×10^5	1.0×10^2	0.028	- primer 1b
	4.4×10^5	3.0×10^2	0.068	- biotinylated dUTP 2
	4.8×10^5	3.0×10^2	0.062	- template 3
10	4.4×10^9	4.0×10^6	0.091	- trypsin
	1.5×10^9	5.5×10^5	0.037	- trypsin, - primer 1b

ϕ_i and ϕ_f denote the number of transformation units (tu) prior [a] and after selection. Yield = ϕ_f / ϕ_i .

15 [c]: + primer 1b, + biotinylated dUTP 2, + template 3 and + trypsin.

Example 9. Selection for disulphide-containing polypeptides.

For the cloning of (poly)-peptide encoding DNA fragments and their display for s
 20 between barnase and p3, the phage fd-3 is constructed (Fig. 5). Phage fd-3 comp
 H102A mutant of barnase N-terminally fused to the p3 gene of phage fd-TET. I
 the codon for the last residue of barnase and the first residue of p3 is the nu
 sequence *CTG CAG GCG GTG CCG CCG CA*. This sequence contains a PstI
 restriction site (in italics) for insertion of DNA fragments flanked by PstI restrict
 25 The sequence further introduces a frame shift between barnase and p3, which
 expression of the correct p3 reading frame in fd-3. Phage particles of phage fd-3
 do not display the infection protein p3 and are non-infectious.

Phage fd-3 is therefore well suited as a cloning vector as vectors without P
 30 inserts after ligation are not propagated during selection. Statistically 1 out of
 DNA inserts in the PstI restriction site will (except for the presence of sto

within the insert) create an open reading frame spanning barnase, the insert itself and p3 and result in infectious phage particles containing p3 in the phage coat. In these recombinant clones barnase is followed by the insert, which is then followed by the amino acid residues AGGAAA before the start of the p3 protein. This AGGAAA peptide should provide enough flexibility in the fusion protein to enable the infectivity function of p3 and the access of the protease to the N-terminal appendices of p3.

Genomic DNA from the *E.coli* strain TG1 is amplified in 30 cycles of a polymerase chain reaction (PCR) with an annealing temperature of 48°C using the oligonucleotide SN6MIX (5'-GAG CCT GCA GAG CTC AGG NNN NNN-3'), which comprises 6 degenerate positions at the extendible 3' end to ensure random priming. In a second step of 30 PCR cycles with an annealing temperature of 52°C primary PCR products are extended by re-amplification with the oligonucleotide XTND (5'-CGT GCG AGC CTG CAG AGC TCA GG-3'). Products with a length of around 150 bp from this reaction are purified from an agarose gel and reamplified in 30 PCR cycles using an annealing temperature of 52°C and the oligonucleotide XTND. These reamplified 150 bp fragments are partially digested with SacI (site indicated in bold in the oligonucleotides) and ligated for dimerisation. Ligated products are reamplified in a further 10 PCR cycles with an annealing temperature of 44°C followed by a 30 PCR cycles with an annealing temperature of 55°C using the oligonucleotide XTND. The annealing temperatures are chosen to discriminate against priming of the oligonucleotide in the middle of the dimerised fragments. The reaction product is size purified twice on an agarose gel to remove monomers and oligomers (non-dimers).

The final dimer fraction is amplified by PCR using an annealing temperature of 55°C and the oligonucleotide XTND on a large scale, digested with PstI (site indicated in italics in oligonucleotides) and ligated into the also PstI digested and phosphatased vector fd-3. After electroporation into *E.coli* bacteria a repertoire of 3.6×10^7 recombinants is obtained. Sequence analysis of randomly picked clones reveal the presence of mainly dimeric (11 out of 14) and some monomeric (2 out of 14) DNA inserts.

Reinfection of *E.coli* bacteria with phage produced from the initial population of transformed cells yields, according to sequence analysis of twenty randomly picked clones, a library of infectious phages containing almost exclusively barnase-(in-frame, no-stop-dimer-insert)-p3 fusions. Non-infectious phages arising from vector without insert and from vector with out-of-frame or stop-codon containing inserts are not propagated in the infection step. The vector fd-3 is therefore suitable to create a repertoire of polypeptides randomly generated through dimerisation of DNA fragments from the *E.coli* genome.

This repertoire of polypeptides displayed as an inserted fusion between barnase and p3 on phage fd is subjected to proteolytic digestion with trypsin and thermolysin alone or a mixture of both (1 ng/μl each) in TBS-Ca buffer (25 mM Tris, 137 mM NaCl, 1mM CaCl₂, pH 7.4). After proteolysis phage is captured with biotinylated barstar bound to a Streptavidin coated microtitre well plate and eluted at pH 2.0. Phage is neutralised to pH 7 and propagated through reinfection of the *E.coli* cells and selected for a second round as before. All steps are performed in the absence of any reducing agent like dithiothreitol (DTT) or β-mercaptoethanol, which would reduce and thereby cleave any potential disulphide bonds formed between cysteine sidechains within selected polypeptides.

Randomly picked phages, which are eluted after the first and second round of proteolytic selection, are analysed for binding to barstar after incubation with a mixture of trypsin and thermolysin under the conditions of the selection (Table 9). Their sequence is determined (Table 9). 17 out of 18 analysed clones treated with a mixture of trypsin and thermolysin during selection are found to bind biotinylated barstar after incubation with trypsin and thermolysin. 5 out of 8 analysed clones treated with trypsin during selection bound biotinylated barstar after incubation with trypsin and thermolysin, whilst 9 of 14 analysed clones treated with thermolysin are found to bind. No randomly picked clones not treated with a protease during selection bind biotinylated barstar after incubation with trypsin and thermolysin. Binding to biotinylated barstar shows that the polypeptide insertion between barnase and p3 on the phage retains its overall covalent integrity and therefore keeps the N-terminal tag

(barnase) and the infection protein p3 (and thereby the phage particle as a whole) covalently linked.

However, the possibility of proteolytic digestion of the peptide backbone can not be excluded, as the inserts may also be kept covalently linked through bonds between sidechain groups like the SH₂ groups of cysteines. Sequence analysis of the selected clones reveal that 19 of 25 (76%) resistant clones contain two or more cysteine residues.

To analyse the role of possible disulphide bonds in the polypeptide inserts, 13 of the selected phages are analysed for binding to biotinylated barstar (and thereby for a covalent linkage of barnase and p3 through the insert) after treatment with trypsin and thermolysin followed by a wash with 20 mM DTT before detection of bound phage. 2 phage clones, which bind barstar after protease treatment without DTT wash and contain no cysteines, are unaffected by the DTT treatment. Another 2 phage clones, which bind barstar after protease treatment without DTT and contain two or more cysteines, are also observed to bind barstar after protease and DTT treatment. This suggests that their inserts are protected from proteolysis of their peptide backbone despite the presence of principle substrate sites for the proteases.

These inserts are therefore protected from proteolytic attack due to a conformational restraint of their peptide backbone. However, 9 phage clones which bind barstar after protease treatment without DTT wash and contain two or more cysteines are observed not to bind barstar after protease and DTT treatment. This suggests that their inserts are proteolytically cleaved in their peptide backbone, but are held together by disulphide bonds between cysteine sidechains in the absence of the reducing agent DTT. These inserts are therefore not protected from proteolytic attack due to a conformational restraint of their peptide backbone.

Thus, the method of the present invention may be configured to select for cysteine-containing polypeptides, even where the polypeptides would be susceptible to protease

attack since the polypeptides are capable of being held together in the selection step by disulphide bonds.

5 **Table 9. Barstar binding of phages displaying barnase-p3 fusion inserts selected after proteolytic treatment under non-reducing conditions and amino acid sequences of their PstI inserts in vector fd-3.**

Barstar binding (-DTT) after proteolysis of phage with trypsin and thermolysin (2ng/ μ l each) without a 20 mM DTT wash is determined by measurement of a signal that is at
 10 least 60% of the signal for barstar binding of phage without protease treatment. Barstar binding (+DTT) after a 20 mM wash is determined by measurement of a signal that is at least 60% of the signal for barstar binding (-DTT) without a 20 mM DTT wash. Phage
 are randomly picked after one (1x) or two (2x) rounds of proteolysis with trypsin (Tr) or thermolysin (Th). Phages are treated with proteases, captured with biotinylated barstar in
 15 microtitre well plates, and washed with 20 mM DTT where applicable. Bound phage are detected with a horse radish peroxidase conjugated anti-M13 phage antibody (Pharmacia Biotech).

	Phage clone	Proteolytic selection	Barstarbindg		Amino acid sequence		
			-DTT	+DTT	of inserts		
5	TA-1.2	1xTr	yes	no	LQSSGDCVIS	DTCIAGMAEA	AACEEKFSSQ
					NVGLTITVTP	CLSSA	
	TA-2.25	2xTr	yes	no	LQSSGCGSSG	SSINCLPCGA	TSRGTSPLAS
10					GLPSSATIHC	LSSA	
	TA-2.26	2xTr	yes	no	LQSSGDSAGC	KNMTGGRLYA	HTLEAIIPGF
					AVSAPACEPA		
15	TA-2.27	2xTr	yes	yes	LQSSGCVRLK	RTSVNHQPDA	WPEPHLKAAC
					EPA		
	TA-2.30	2xTr	yes	no	LQSSGCGSSG	SSINCLPCGA	TSRGTSPLAS
20					GLPSSATVQC	LSSA	
	TB-1.10	1xTh	yes	yes	LQSSGKIVQA	GANIQDGCIM	HGYCDTDTIV
					GENGHIGLSS	A	
25	TB-1.11	1xTh	yes	yes	no insert, Barnase & p3 in frame		
	TB-2.33	2xTh	yes	no	LQSSGVCVIS	DTCIAGTAEA	AACEEKFSSQ
					NVGHTITETP	CLSSA	
	TB-2.34	2xTh	yes	no	LQSSGCGSSG	SSINCLPCGA	TSRGTSPLAS
					GLPSSATIQ	LSSA	
	TB-2.35	2xTh	yes	no	LQSSGQDSQR	EHASHTAEDD	CEDQTRIHQH
					IREVDFVDTP	QEVDDCRAAL	SSA
	TB-2.37	2xTh	yes	no	LQSSGCVRLK	RTSVNHQPDA	WPEPHLKAAC
					EPA		
	TB-2.38	2xTh	yes	yes	LQSSGVRPA		
	TB-2.39	2xTh	yes	no	LQSSGCGSS	GSSINCLPCGA	TSRGTSPLAS
					GLPSSATIQ	CLSSA	

Example 10. Use of reducing agents to eliminate disulphide-related background.

The repertoire of polypeptides described in Example 9 is digested as before with both trypsin and thermolysin (Example 9) except for an additional washing step here with 50 mM DTT after binding of the proteolytically treated phage to the microtitre well plates coated with Streptavidin-biotinylated barstar. This washing step is designed to wash off phage displaying an N-terminal barnase tag, which is no longer linked to p3 through an intact polypeptide backbone but only through disulphide bonds between cysteine sidechains in the polypeptide inserts between the barnase tag and p3.

12 randomly picked phages eluted after the second round of proteolysis are analysed for stability against a mixture of trypsin and thermolysin under the conditions of the selection and their sequence is determined (Table 10). 10 clones treated with a mixture of trypsin and thermolysin during selection bind biotinylated barstar after incubation with trypsin and thermolysin followed by washing with 50 mM DTT before detection of captured phage. Only one of these clones contain two or more cysteines.

Proteolytic treatment of the phage library followed by a wash with DTT therefore allows the selection of peptide inserts which are protected from proteolysis and which are not held together through disulphide bonds.

Table 10. Barstar binding of phages displaying barnase-p3 fusion inserts selected after proteolytic treatment followed by treatment with 50 mM DTT and amino acid sequences of their PstI inserts in fd-3. Barstar binding (+DTT) after proteolysis of phage with trypsin and thermolysin (2 ng/μl each) followed by a 50 mM DTT is determined by measurement of a signal for Barstar binding (+DTT), which is at least 60% of the signal for barstar binding of phage without protease treatment. Phage are randomly picked after two (2x) rounds of proteolytic with trypsin (Tr) and/or thermolysin (Th). Phages are treated with proteases, captured with biotinylated barstar in microtitre well plates, and washed with 50mM DTT where applicable. Bound phage is detected with a horse radish peroxidase conjugated anti-M13 phage antibody.

	Phage Clone	Proteolytic selection	Barstarbindg +DTT	Amino acid sequence of inserts
15	B2-13	2xTr/Th	yes	LQSSGTEVDR GNQQHDTNDR DFTHTPLSS A
	B2-14	2xTr/Th	yes	LQSSGSVAQG SSASVDVTAT NAVLSADSL SLGGGEPA
	B2-22	2xTr/Th	yes	LQSSGGAVAV TPGPVLSSA
20	B2-23	2xTr/Th	yes	LQSSGHCGRK PVLCTHTA
	B2-15	2xTr/Th	yes	LQSSGVRPA
	B2-17	2xTr/Th	yes	no insert, Barnase & p3 in frame
	B2-20,21	2xTr/Th	yes	no insert, Barnase & p3 in frame
	B2-16,24	2xTr/Th	yes	LQSSGEPAPA HEAKPTEAPV AKAEAKPETP AHLSSA
25	B2-18	2xTr/Th	no	LQSSGCVRLK RTSVNHQPD A WPEPHLKAAC EPA
	B2-19	2xTr/Th	no	LQSSGVVDWA KMREIADSIG AYLFVDMAHV AALSSA

Brief description of Figures

Figure 1. Cleavage of phages with protease sites. Phages were prepared by rescue with KM13 (pHEN1, A + B), or with VCSM13 (pK1, C + D). Uncleaved (A + C) or
 5 cleaved with trypsin (B + D). 5 μ l, 2.5 μ l and 1 μ l phages were loaded as indicated. Molecular weight markers are in kD.

Figure 2. The phagemid vectors pK1 and pK2. These vectors contain a protease cleavable sequence between D2 and D3 of the phage p3 protein. In pK1, D2 + D3 are
 10 in frame; in pK2, D3 is out of frame.

Figure 3. Binding of phage-barnase to barstar. Phage displaying different fusion protein are incubated with biotinoylated barstar captured on streptavidin-coated plate and detected by ELISA. a) barnase mutant A, b) barnase mutant B, c) villin , d) no phage.
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Figure 4. Temperature denaturation of phage fusion proteins. Phagemids were rescued with KM13, infectivity (TU/ml) shown after incubation and cleavage with trypsin at given temperatures. Fusion with villin subdomain (triangles), barnase mutant A (diamonds), barnase mutant B (squares), pHEN1-chloramphenicol resistant (circles).
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Figure 5. The fd vector fd-3. The gene for the H102A mutant of Barnase is introduced by subcloning into fd-DOG [43] after PCR amplification with suitable oligonucleotides using the restriction sites ApaLI (at the Barnase 5' end) and NotI to create fd-3.

Figure 6. Map of phagemid vector used for display of Stoffel fragment on the surface of phage. The asterisk shows sequences that are randomised at least partially in libraries I and II.
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Claims

1. A method for the selection of a virus comprising the steps of:
 - (a) providing a virus encoding and displaying a fusion polypeptide, said fusion polypeptide comprising a heterologous polypeptide inserted into the sequence of a viral coat protein polypeptide, wherein said virus comprises a cleavable site located within a displayed polypeptide;
 - (b) exposing the virus to a cleaving agent;
 - (c) propagating the virus comprising intact fusion protein.
2. A method according to claim 1 in wherein the cleavage site is located within the fusion polypeptide.
3. A method according to claim 2 wherein after cleavage, the virus comprising uncleaved fusion polypeptide is separated from virus comprising cleaved fusion polypeptide.
4. A method according to claim 1 wherein cleavage impairs the ability of the polypeptide comprising the cleavage site to mediate the infection of the virus.
5. A method according to claim 1, wherein the virus encodes a repertoire of sequences.
6. A method according to claim 5, wherein the repertoire of sequences encodes the displayed heterologous peptide or protein.
7. A method according to any one of claims 5 or 6 in which the cleavable site is comprised within the repertoire of sequences.
8. A method according to claim 1, wherein a virus that is resistant to cleavage is propagated by infection.
9. A method according to claim 8 in which a virus which is resistant to cleavage displays a folded protein or polypeptide.

[illegible]

Figure 1.

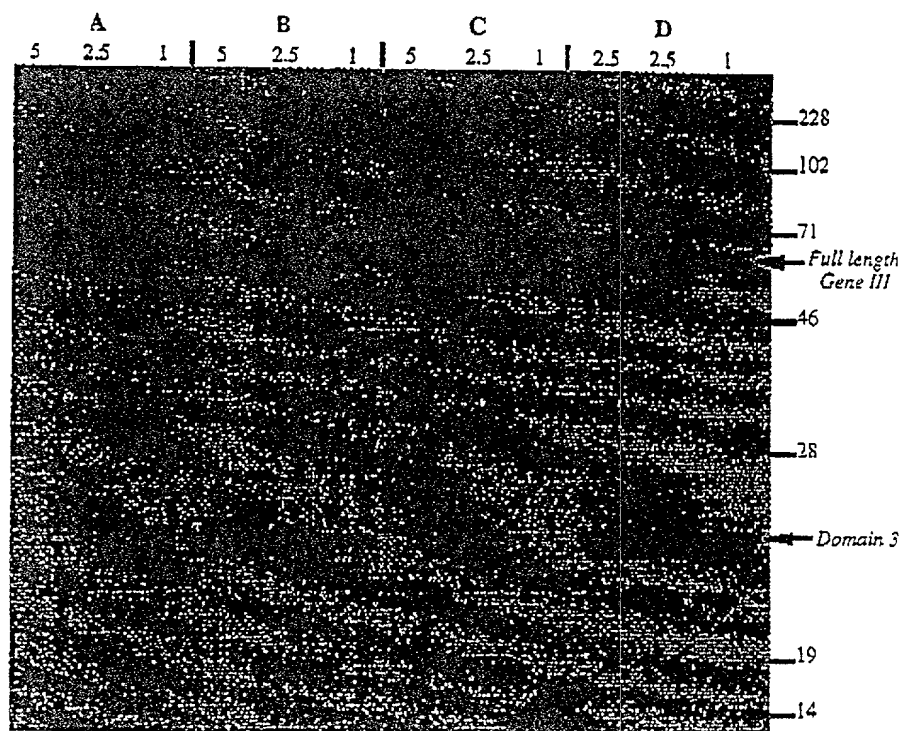


Figure 2.

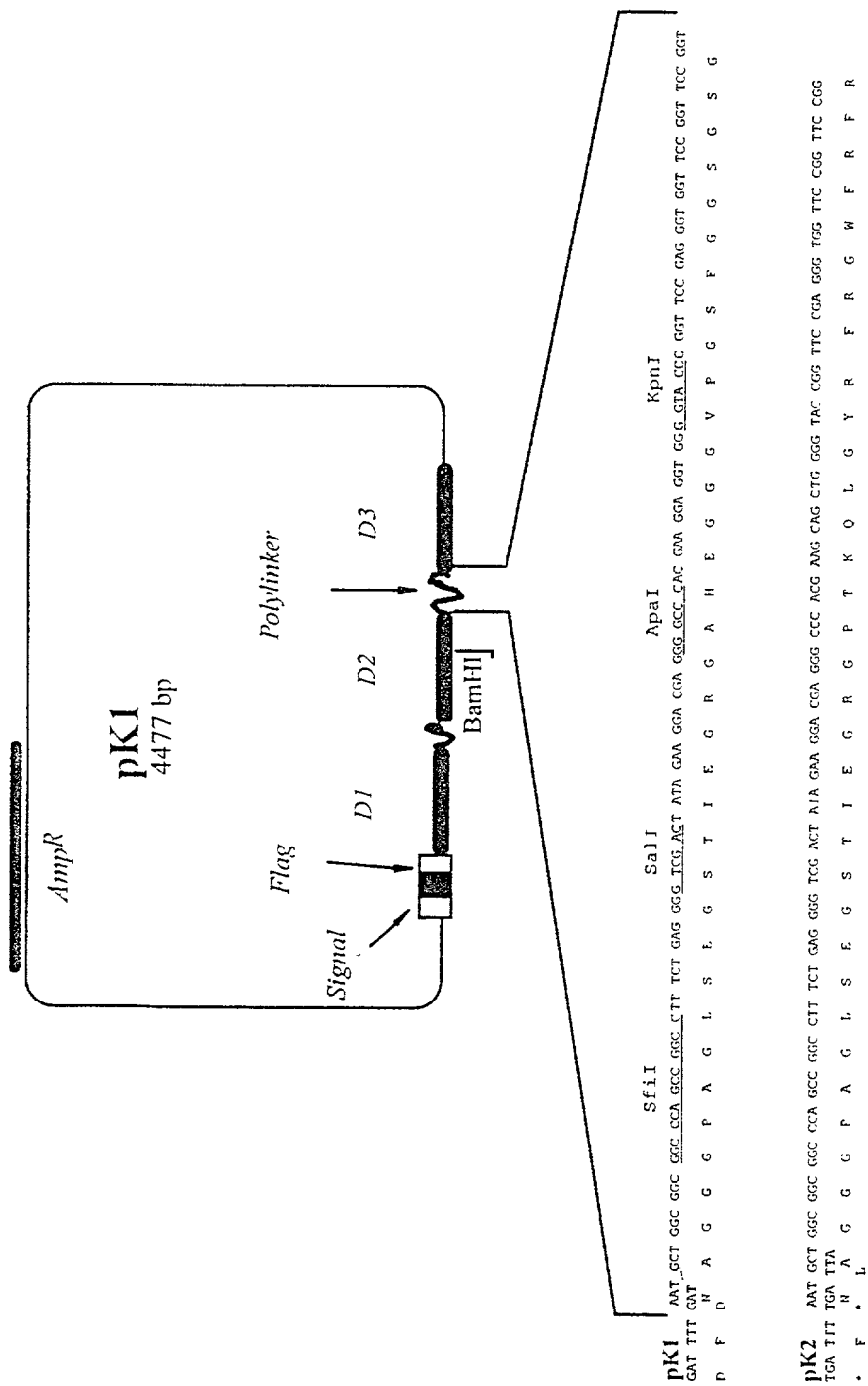


Figure 3.

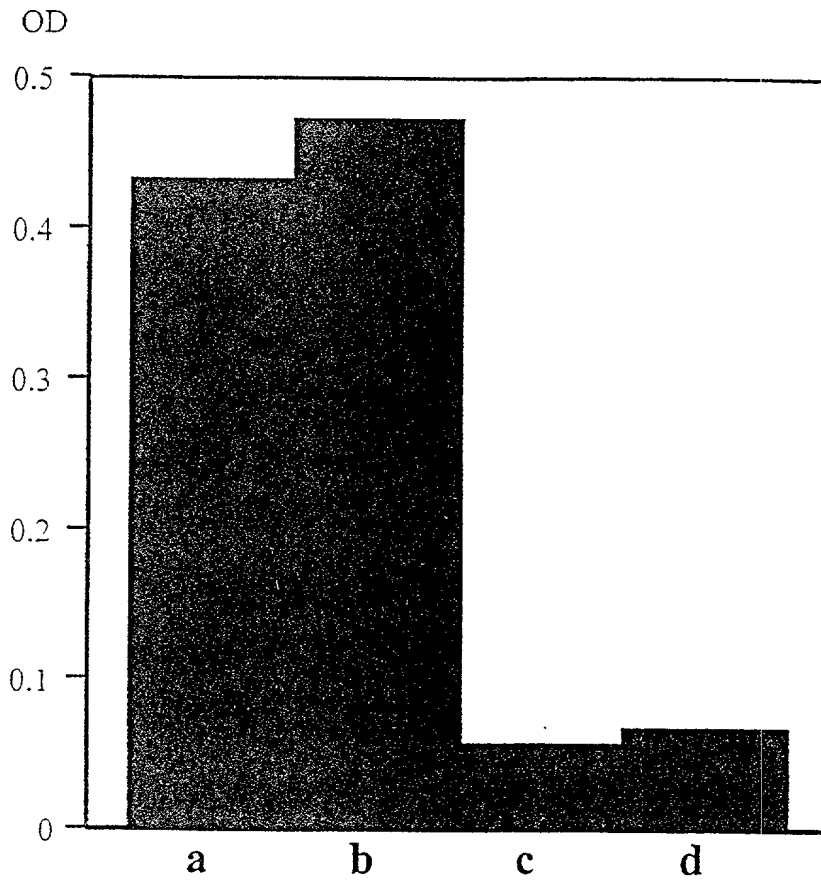


Figure 4.

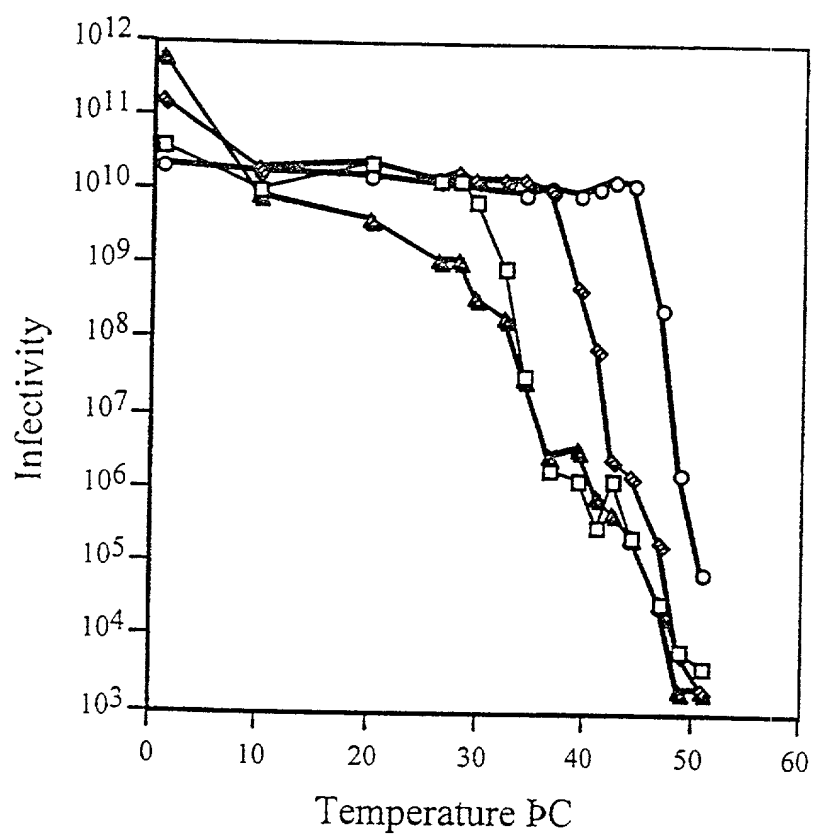


Figure 5.

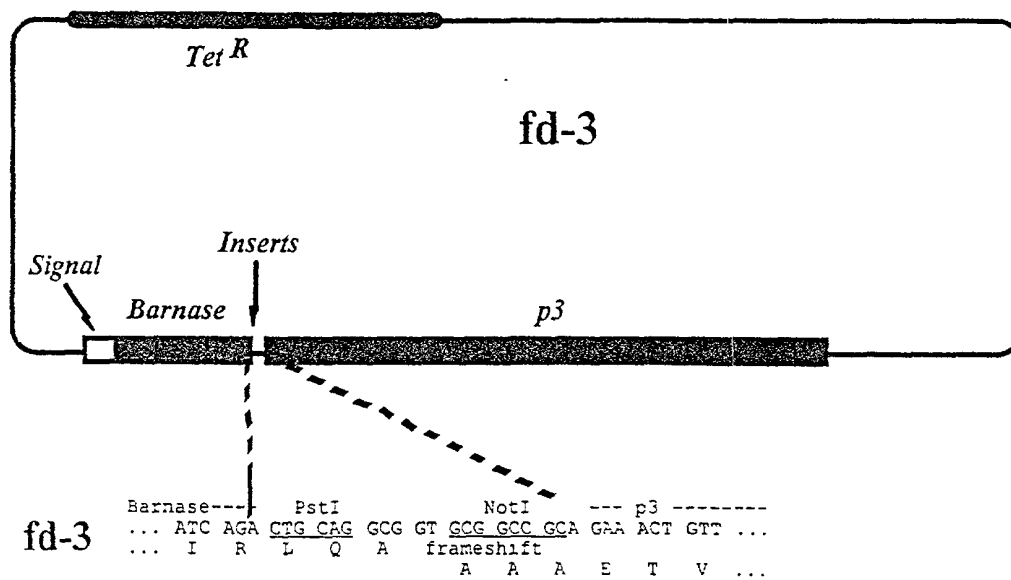


Figure 6.

